

Inflammation and extracellular proteinases



BHI

Bristol Heart Institute

NHS

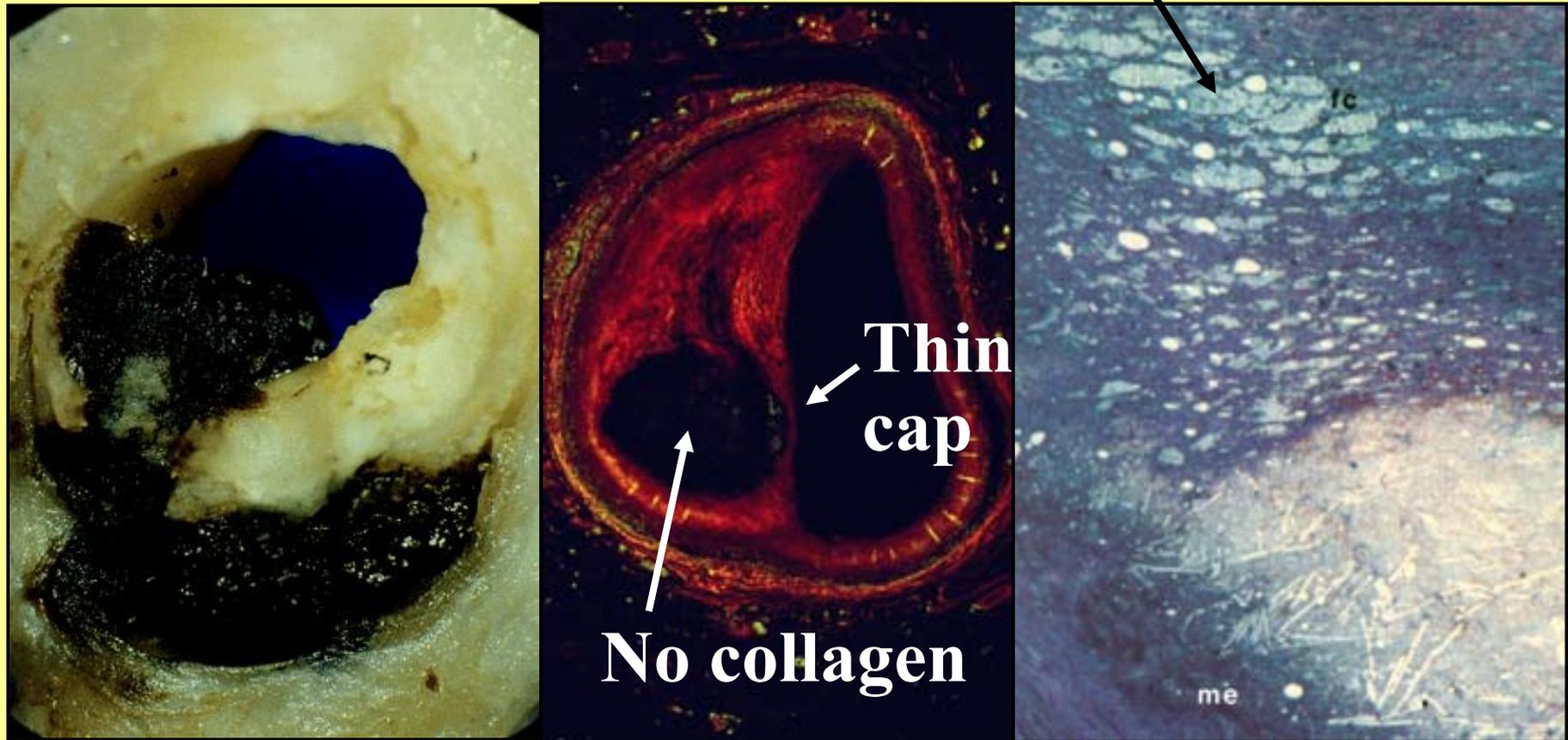
*National Institute for
Health Research*



**British Heart
Foundation**

Plaque rupture 75% of MI (heart attack)

Foamy macrophages

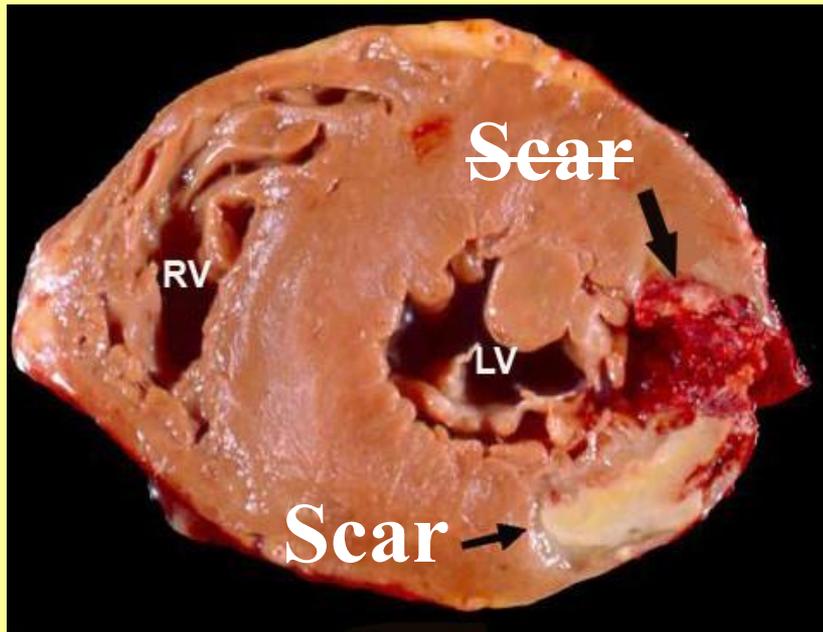


Thin fibrous cap ($<65 \mu\text{m}$), large lipid core, no collagen, **macrophages**, T-cells

Davies, M Circulation 1996; Falk, E JACC 2006;47:C7-12

Long-term consequences of MI

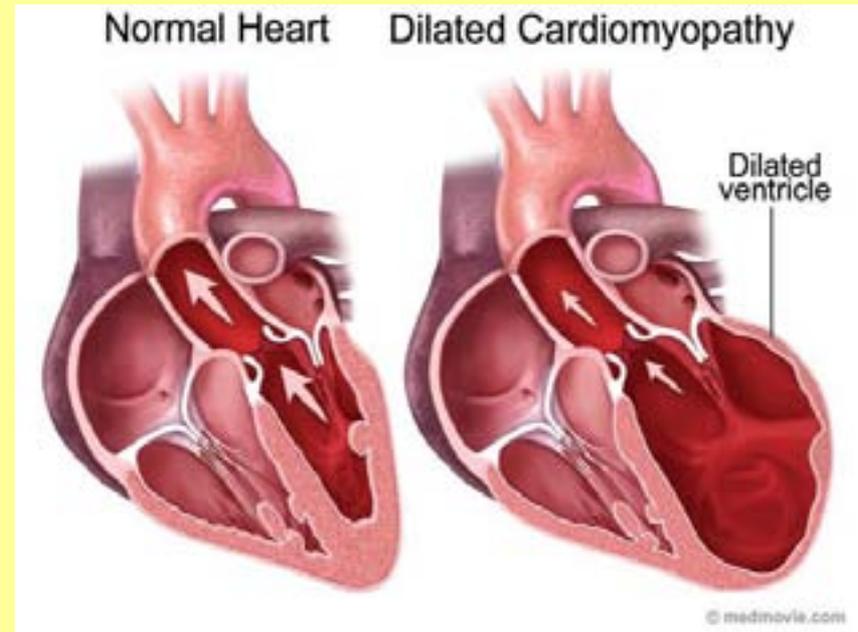
Cardiac rupture



www.readcube.com

**Too little
Inflammation/ repair**

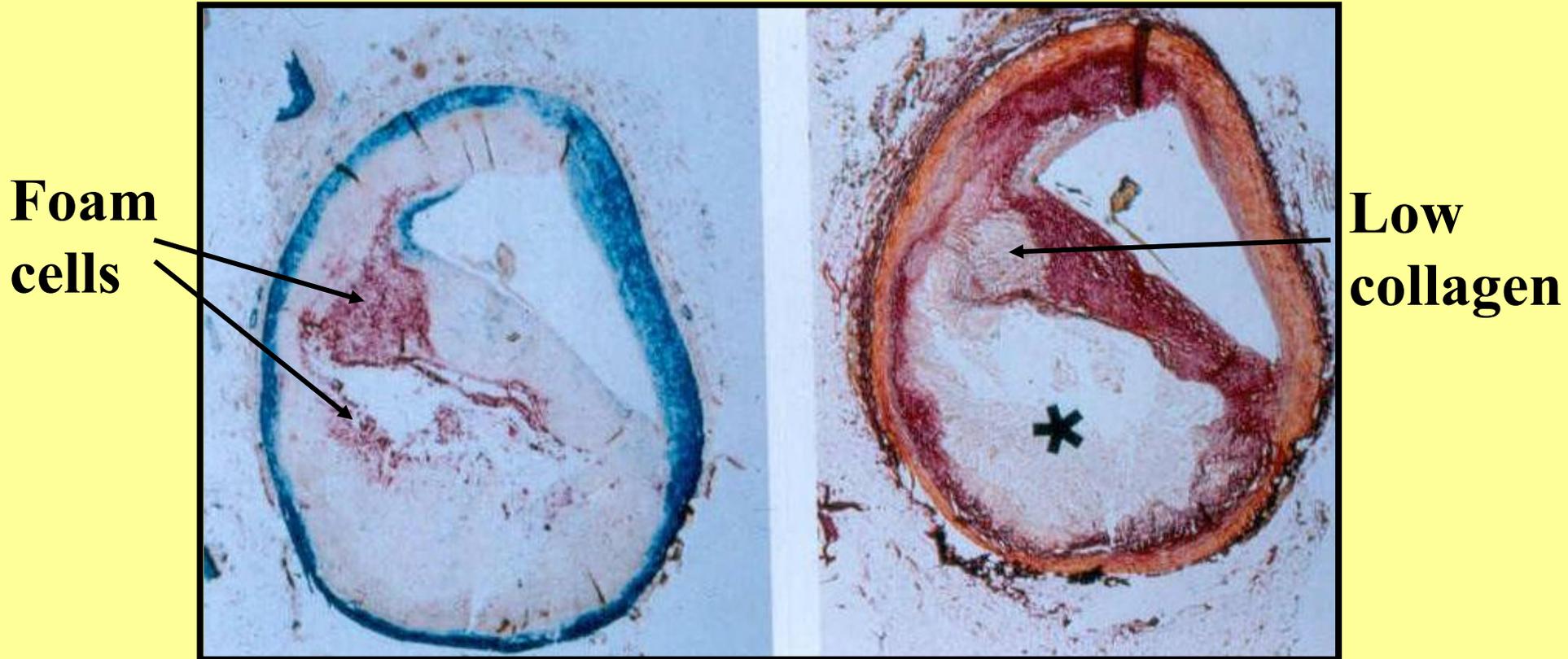
Heart failure



stanfordhospital.org

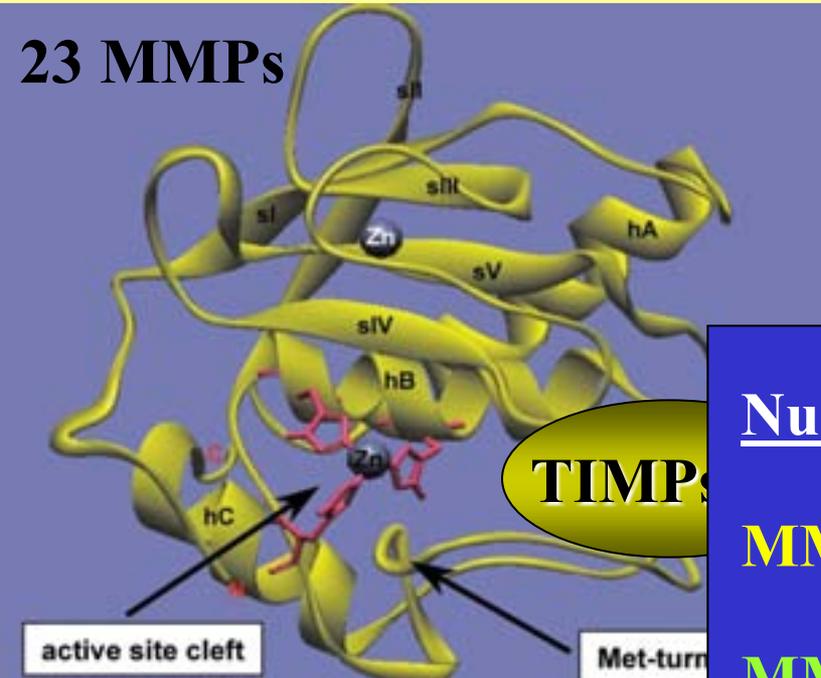
Too much inflammation

Foam cells make proteases that cause plaque vulnerability



Metalloproteinases and TIMPs

Many non-matrix substrates



Numbers

MMP class

MMPs 1,8,13

Interstitial collagenases

MMPs (2),9

Gelatinases (A),B

MMPs 3,7,10,11

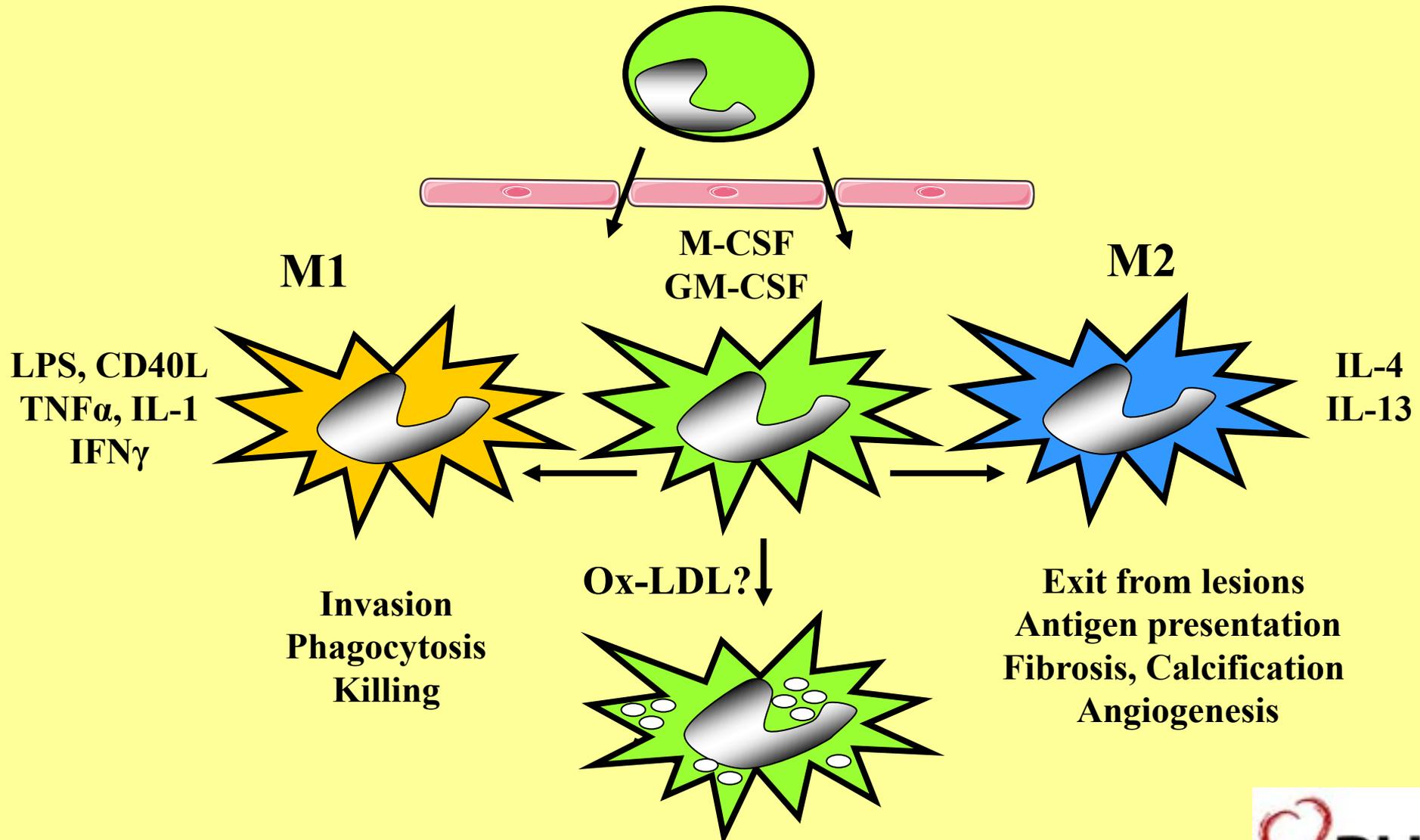
Stromelysins

MMP 12,19

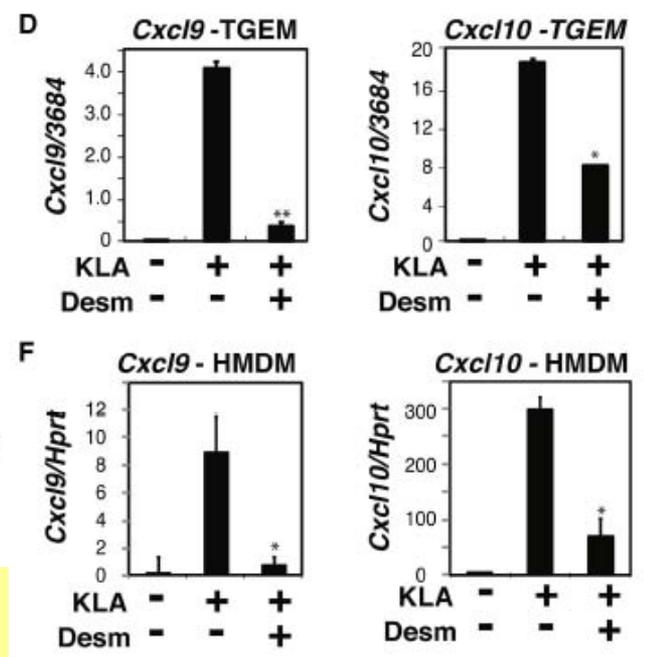
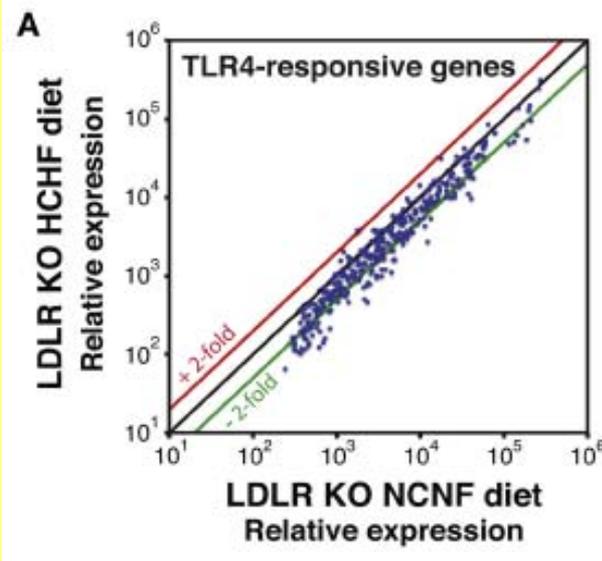
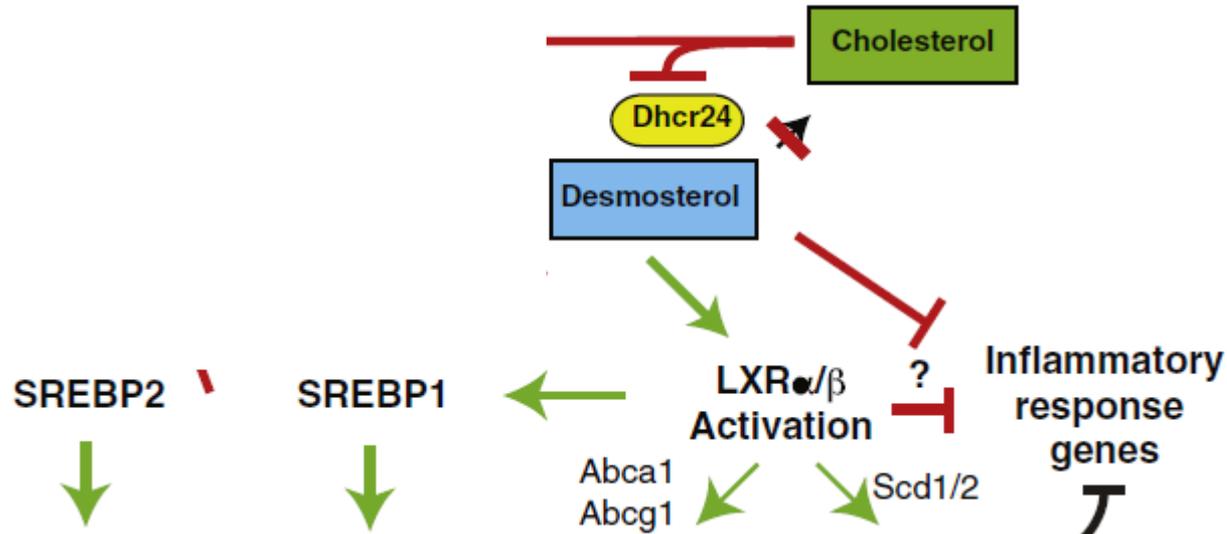
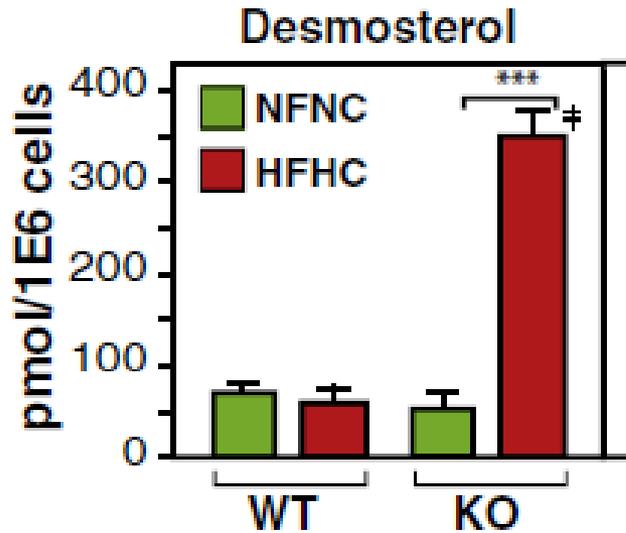
Metalloelastase +

MMPs 14-7,23,25 Membrane-type MMPs

Inflammation and macrophage diversity



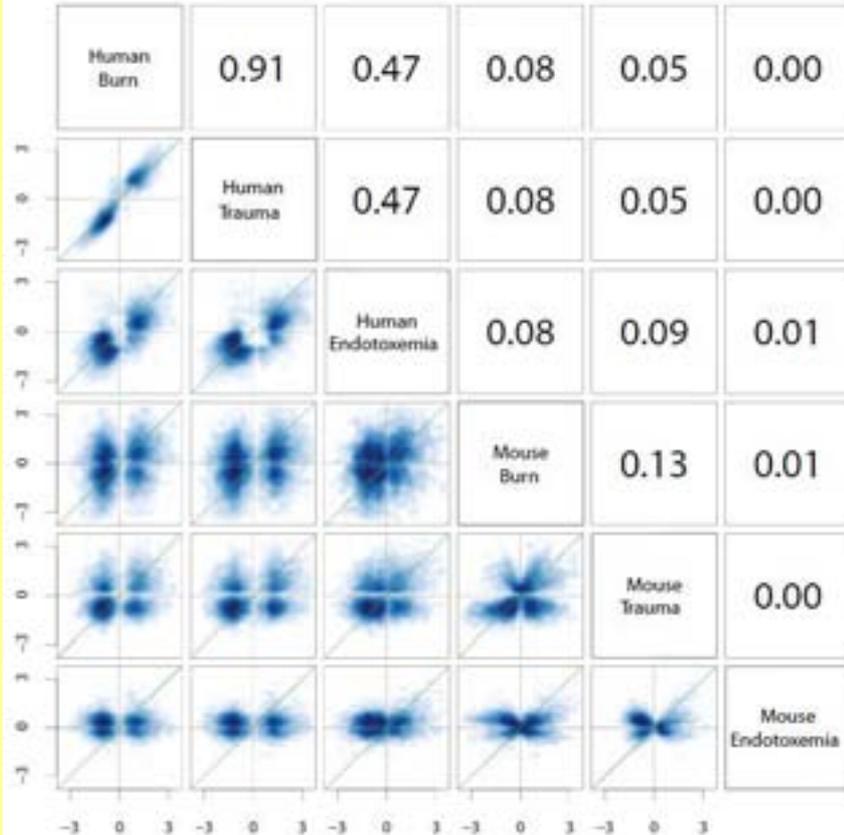
Effect of foam cell formation



Genomic responses in mouse models poorly mimic human inflammatory diseases

Junhee Seok^{a,1}, H. Shaw Warren^{b,1}, Alex G. Cuenca^{c,1}, Michael N. Mindrinos^a, Henry V. Baker^c, Weihong Xu^a, Daniel R. Richards^d, Grace P. McDonald-Smith^e, Hong Gao^a, Laura Hennessy^f, Celeste C. Finnerty^g, Cecilia M. López^c, Shari Honari^f, Ernest E. Moore^h, Joseph P. Mineiⁱ, Joseph Cuschieriⁱ, Paul E. Bankey^k, Jeffrey L. Johnson^l, Jason Sperry^l, Avery B. Nathens^m, Timothy R. Billiar^l, Michael A. Westⁿ, Marc G. Jeschke^o, Matthew B. Kleinⁱ, Richard L. Gamelli^p, Nicole S. Gibran^l, Bernard H. Brownstein^q, Carol Miller-Graziano^h, Steve E. Calvano^c, Philip H. Mason^e, J. Perren Cobb^s, Laurence G. Rahme^s, Stephen F. Lowry^{r,2}, Ronald V. Maier^l, Lyle L. Moldawer^c, David N. Herndon^g, Ronald W. Davis^{a,3}, Wenzhong Xiao^{a,t,3}, Ronald G. Tompkins^{c,3}, and the Inflammation and Host Response to Injury, Large Scale Collaborative Research Program⁴

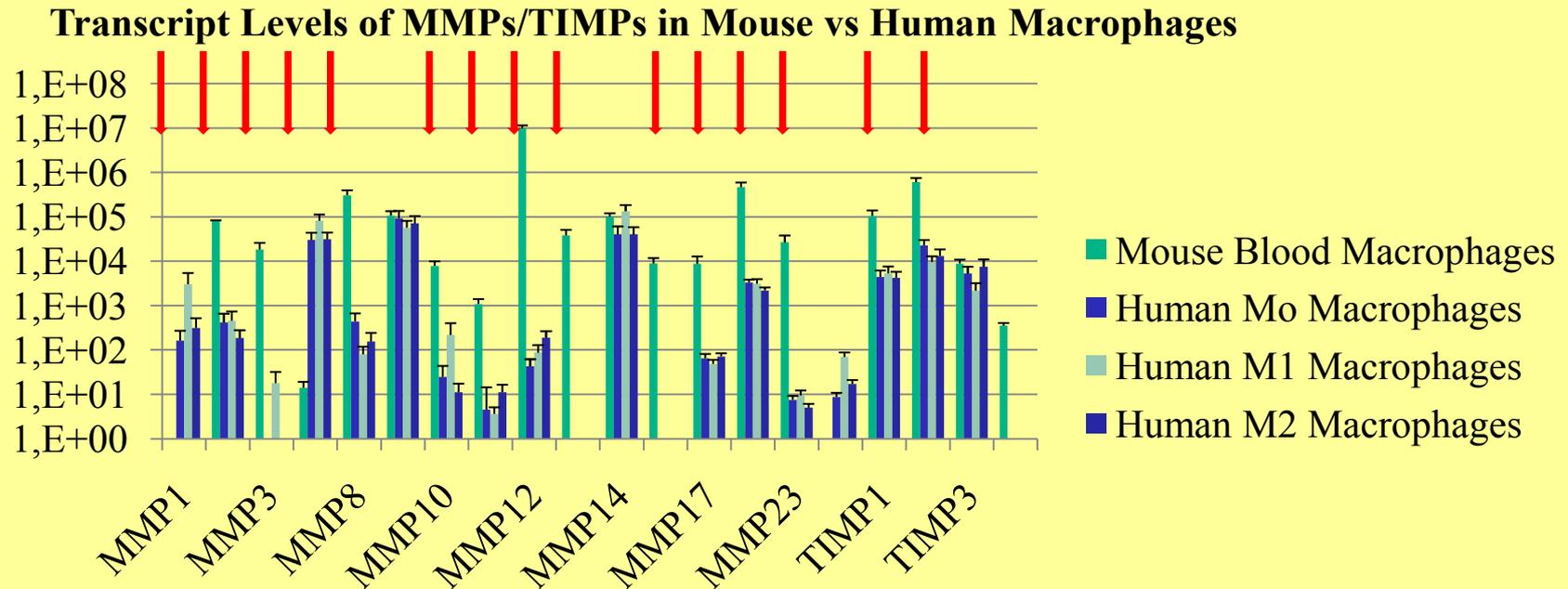
PNAS | February 26, 2013 | vol. 110 | no. 9 | 3507–3512



Are all mouse models of atherosclerosis Dinosaurs?

Many biochemical differences

- HDL rather than LDL
- SAA rather than CRP
- MMP-13 rather than MMP-1



- Many MMP/TIMPs showed 10-10,000 fold Δ

Macrophage phenotypes – the future



Follow the pathways!



David Hockney

MCSF, GMCSF and OxLDL

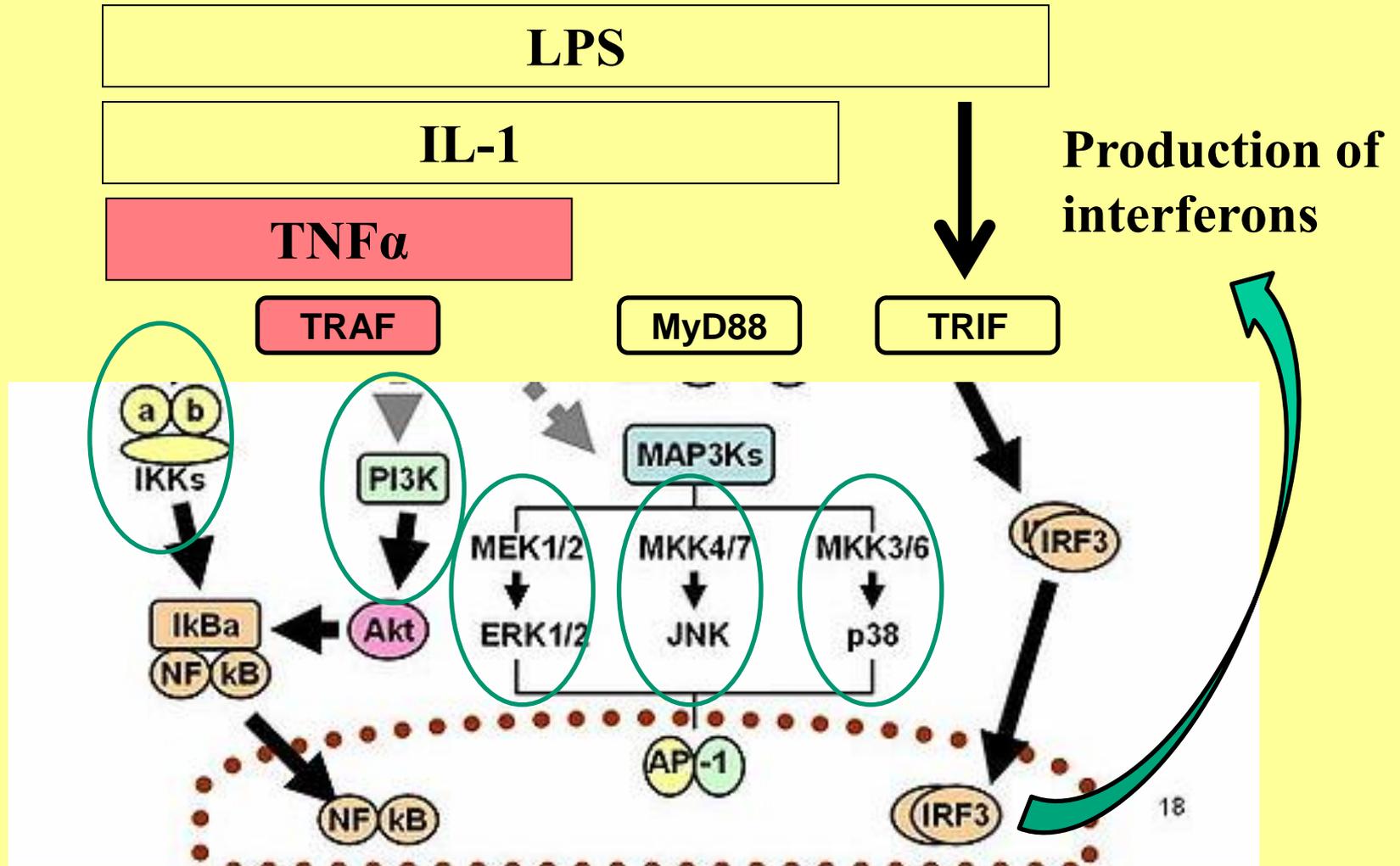
Old name	M-Mac	GM-Mac	Foam cell
Proposed name	MM(-CSF)	MGM(-CSF)	FM(OxLDL)
	CSFR1/CD115	CSFR2/CD116	Scavenger Rs
Signal 1	ERKs, PI-3K	ERKs, PI-3K	Free cholesterol, desmosterol
TFs	Pu.1, Egr-1 AP-1	Pu.1, Egr-1 AP-1, STAT-5	LXRα,β SREBPs
Markers	CD206	CD206	CD11c, Foamy appearance
Proposed marker		Nuclear STAT-5P	Adipophilin

1

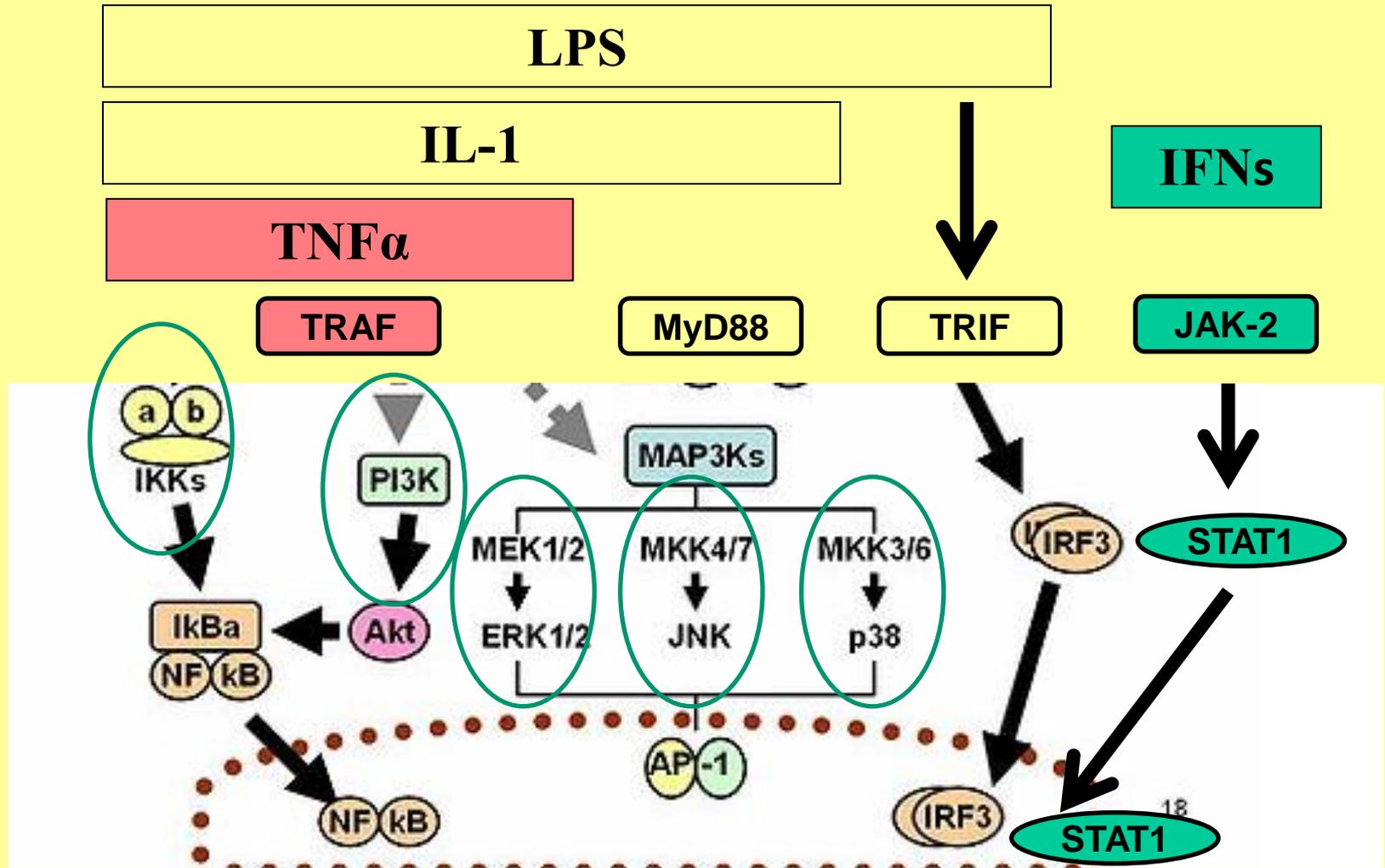
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Classical activation pathways



Classical activation pathways



Classically-activated (M1) phenotypes

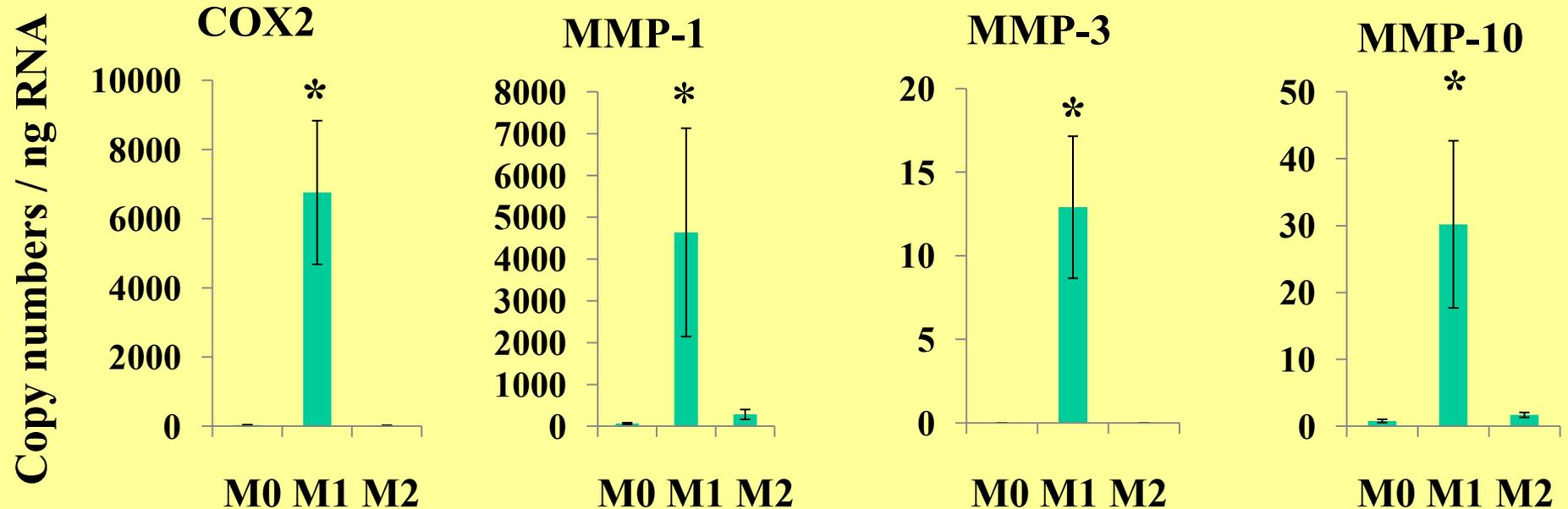
Proposed name	MTNF α	MCD30L /CD40L /Ox40L	MIL-1	MLPS (TLL)	MIFN γ	(M1) MLPS +IFN γ
Receptor	TNFRSF		IL1R1	TLR-4 (TLRs)	IFN γ R1	
Signal 1	TRAFs		Myd88	Myd88 IRF-3	Jak1/2, IRF-3	All
Signal 2	ERKs, PI3K, PKC, IKK2				STAT-1	All
TFs	AP-1, NF- κ B				STAT-1	All
Markers?	I κ B, COX-2, iNOS				IL12, TLR4, CD14	Both
Proposed marker	Nuclear NF- κ B				Nuclear STAT1-P	Both

4

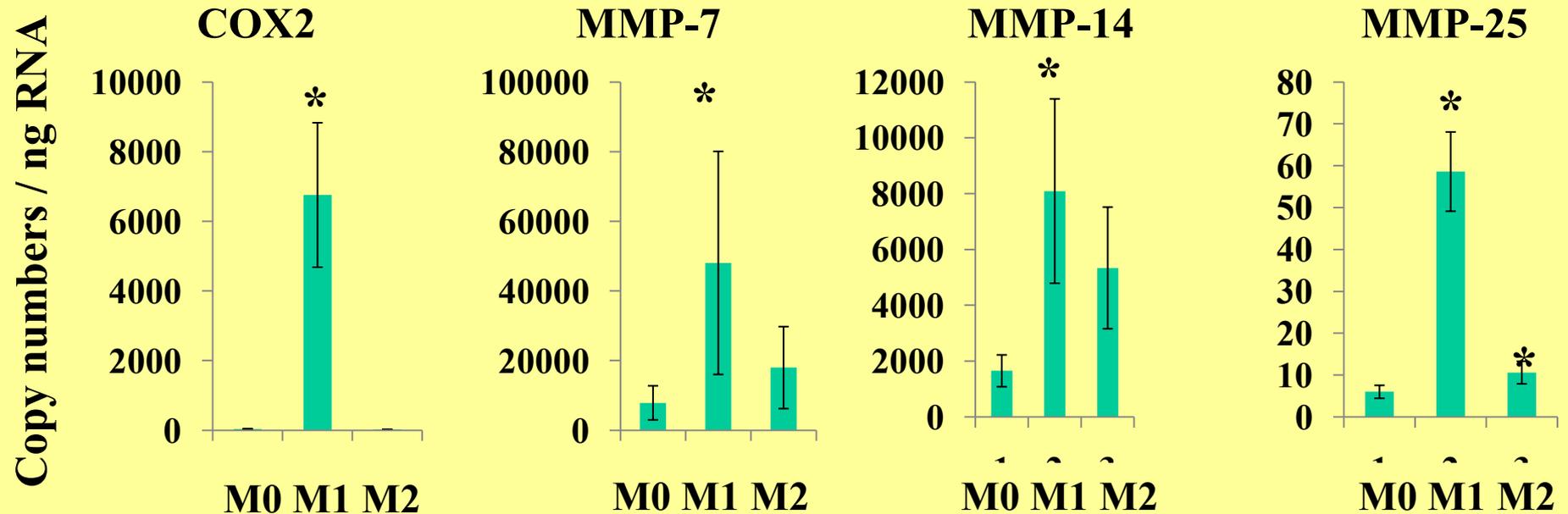
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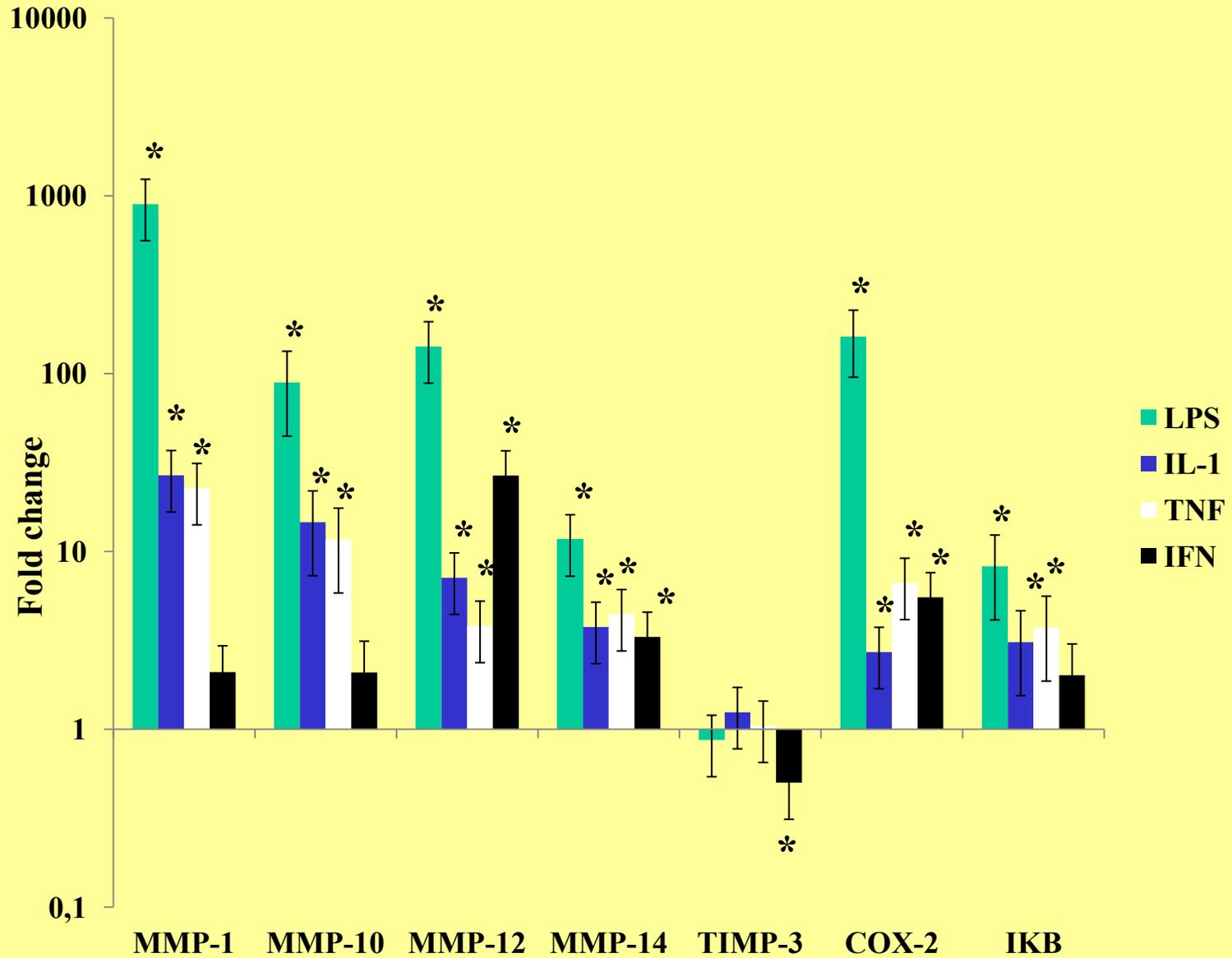
Human Foam cells stimulated by LPS + IFN γ in vitro (M1)



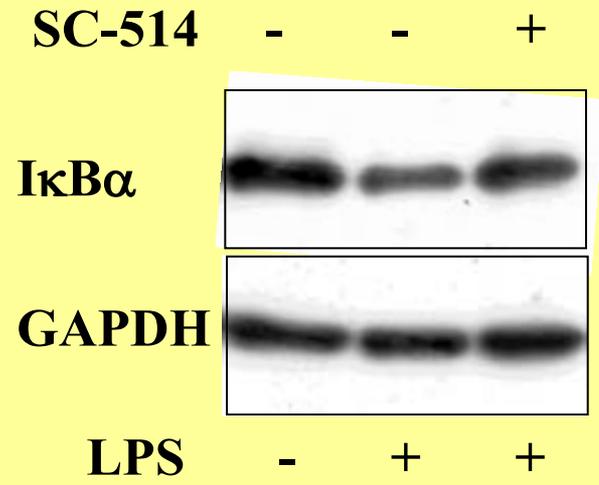
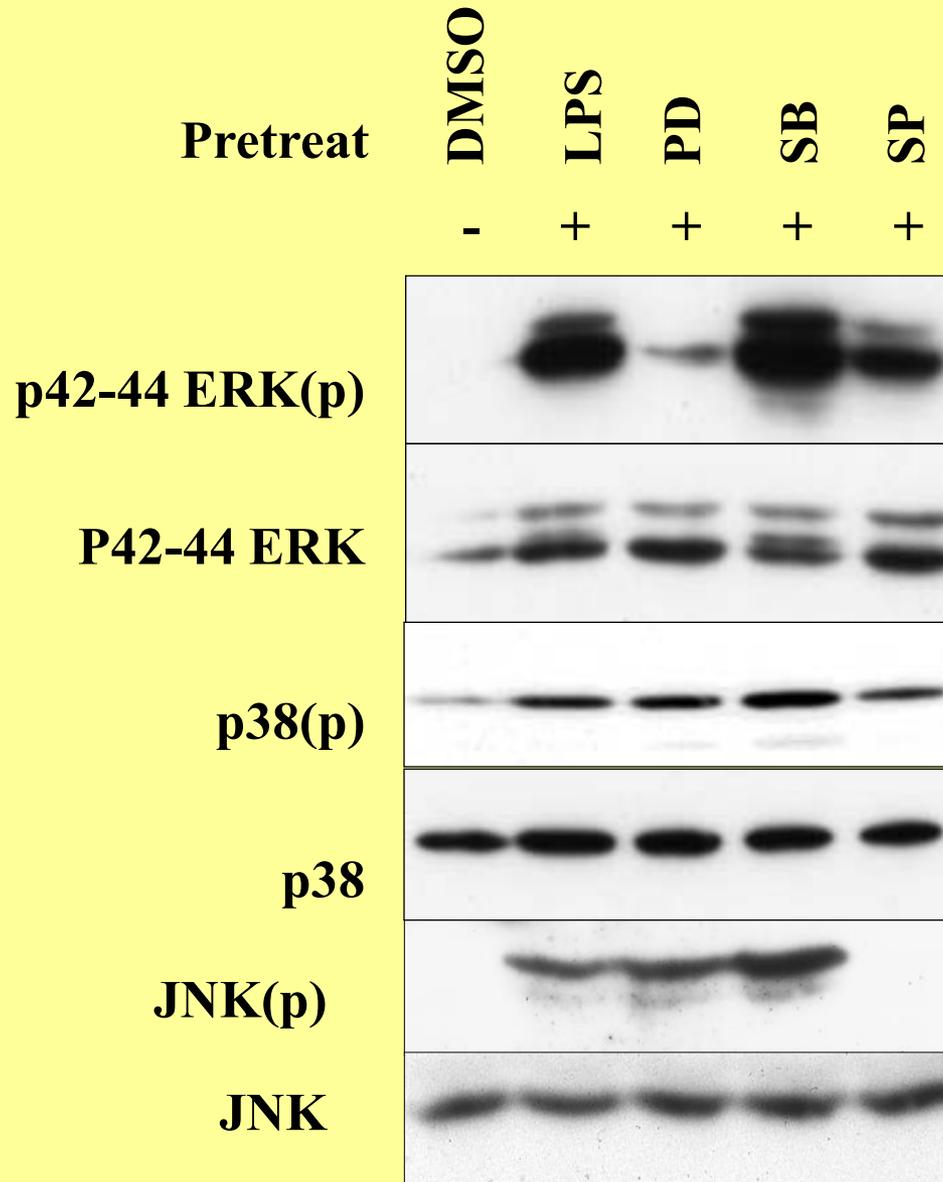
Human foam cells stimulated by LPS + IFN γ (M1) in vitro



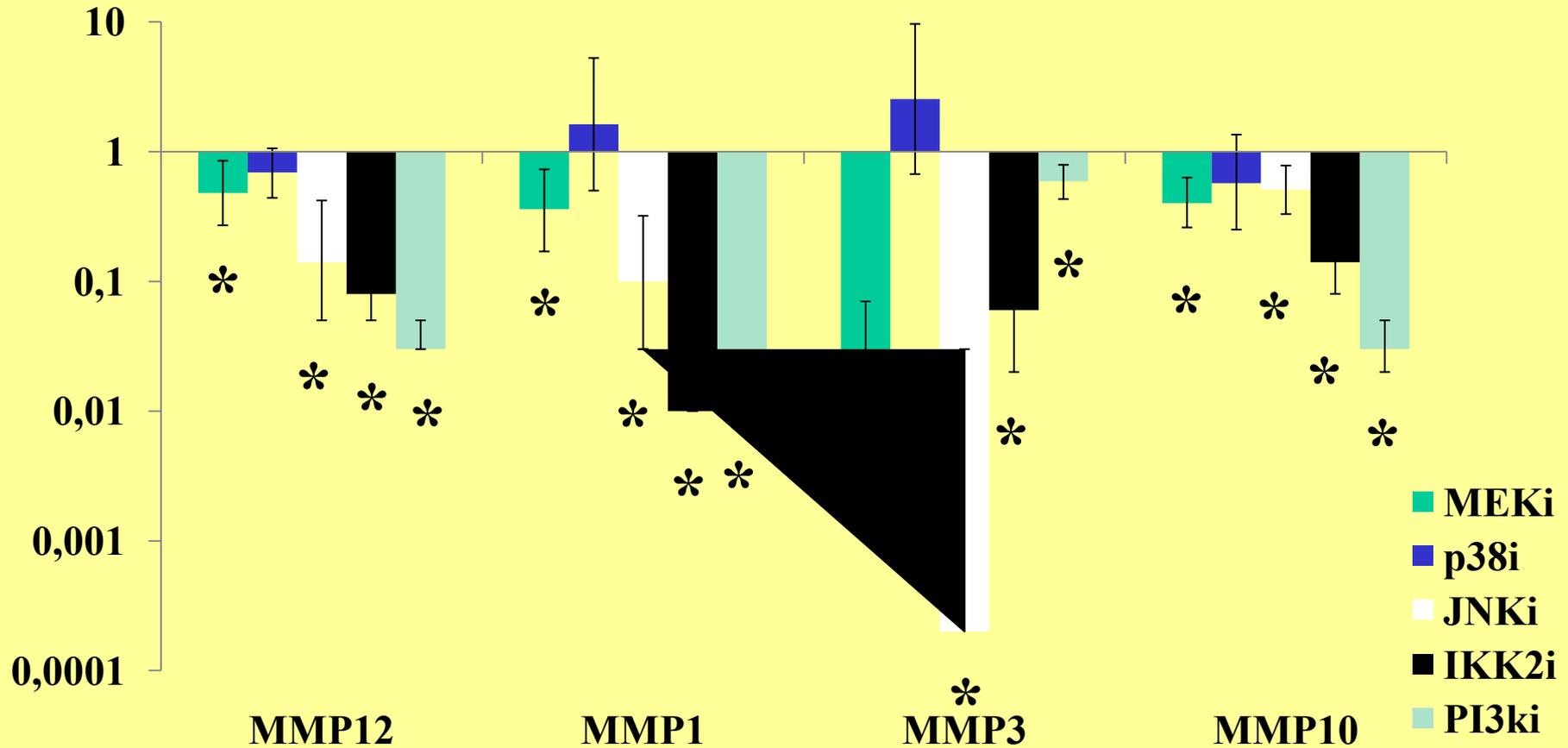
Effects of classical activators



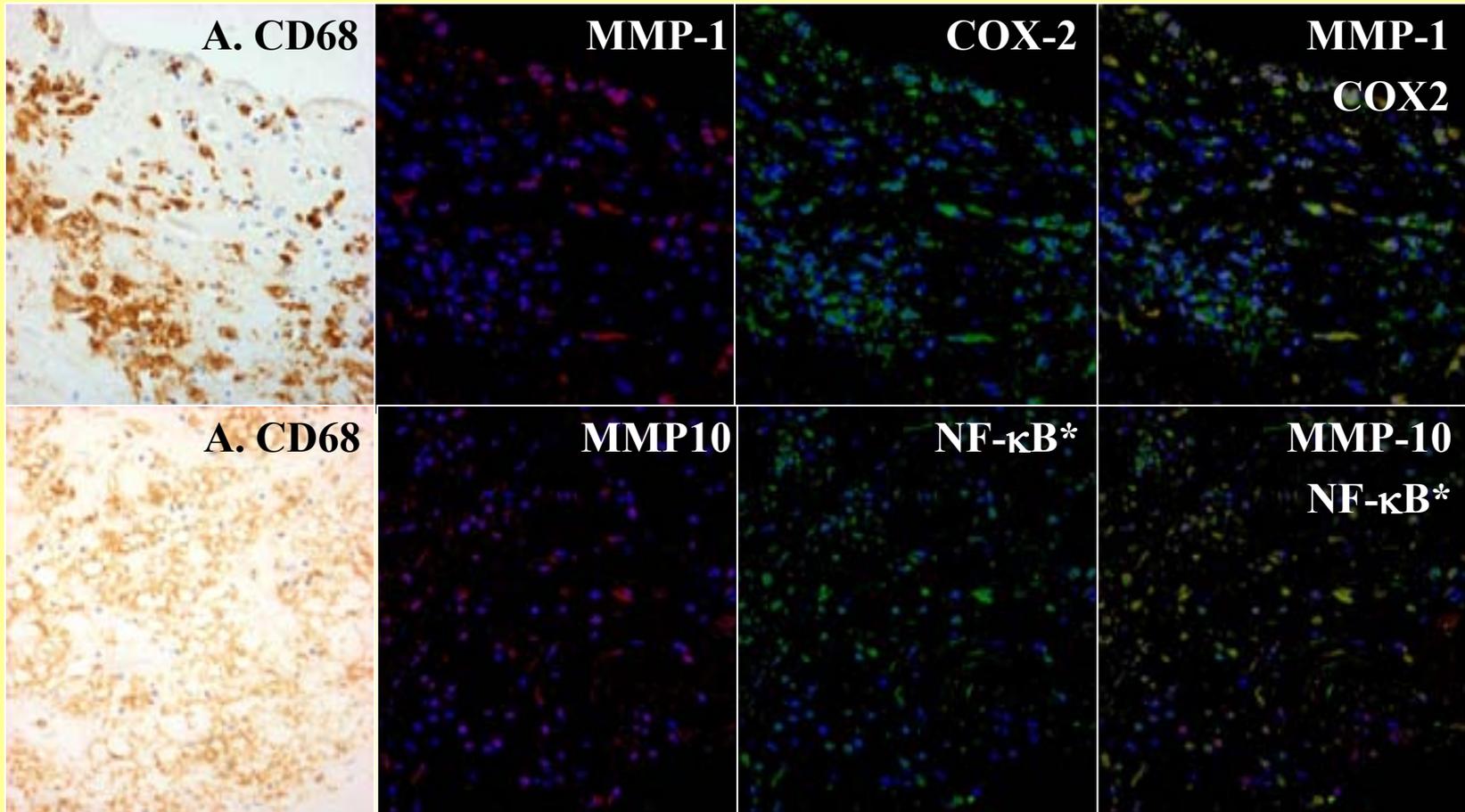
LPS on MAP and IκB kinases



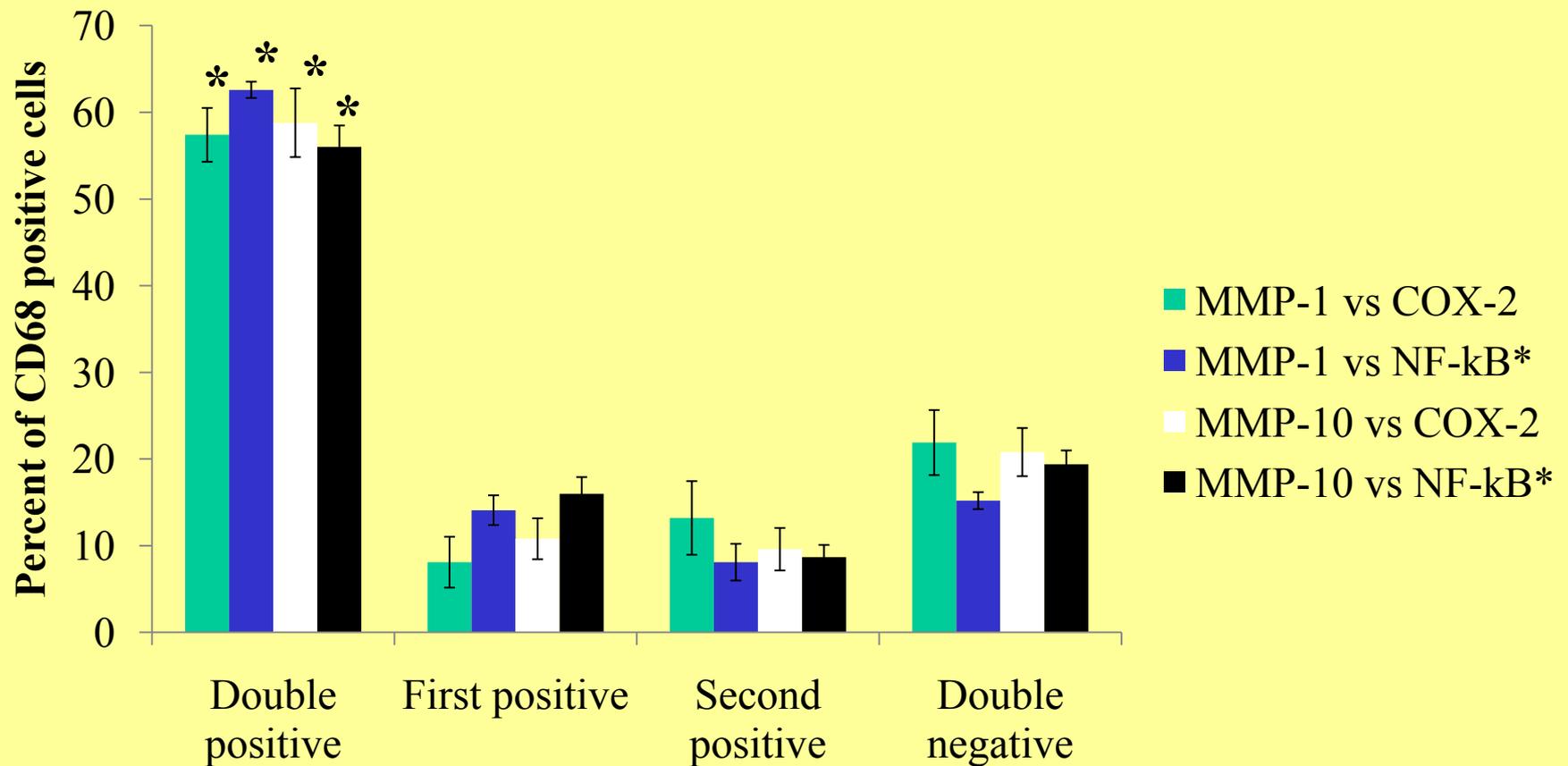
Effect of inhibitors on LPS



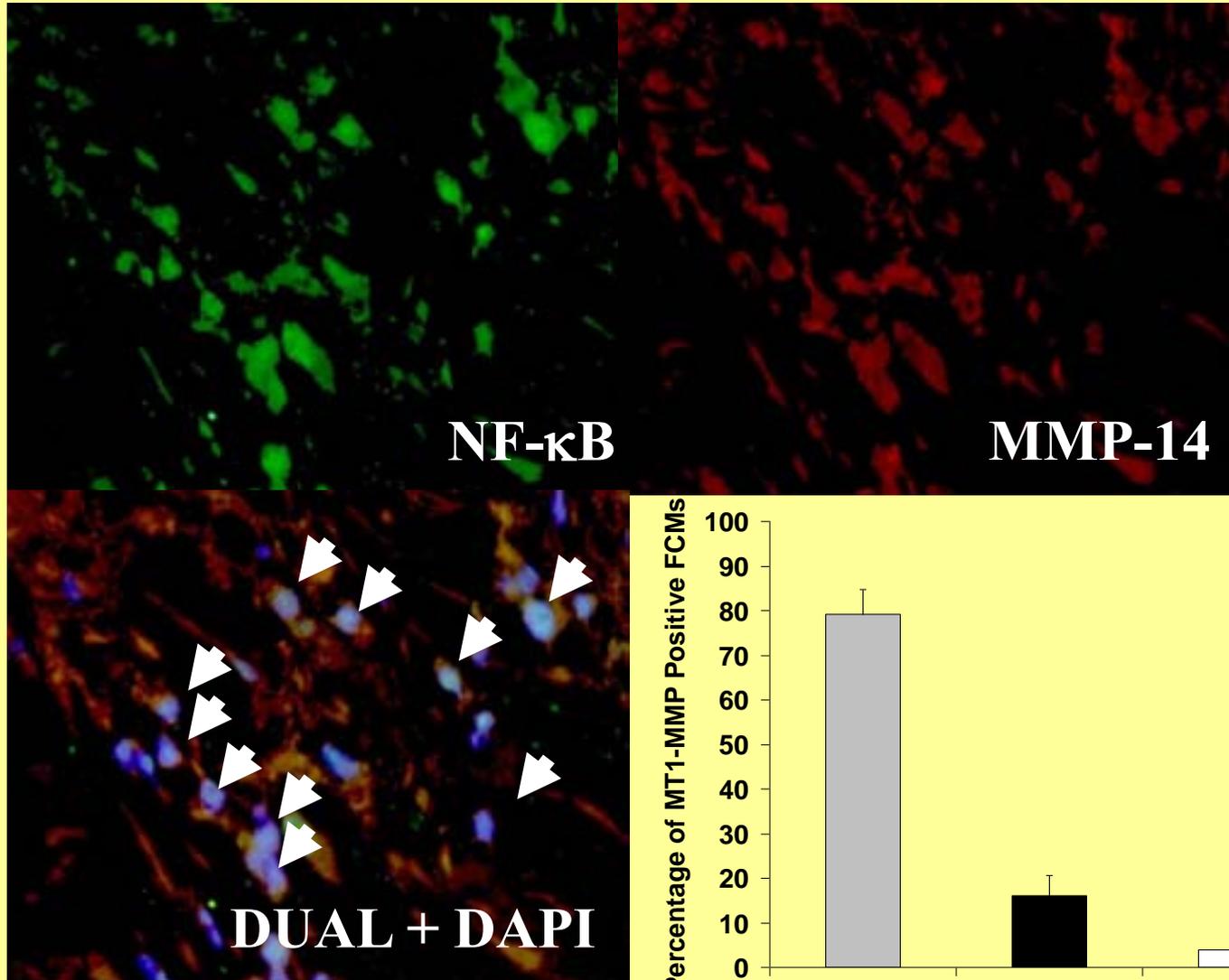
Co-localisation in human coronary macrophages



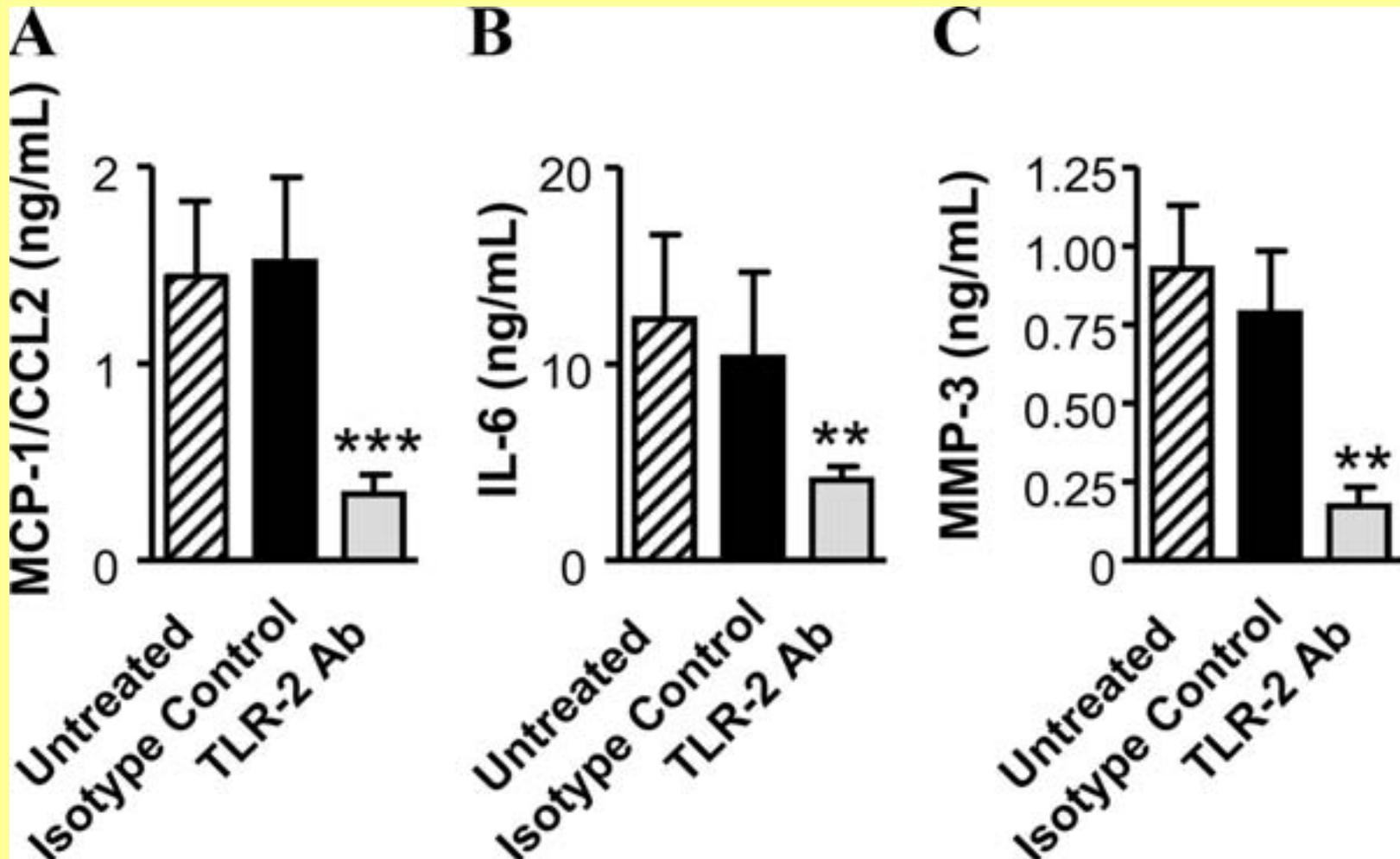
Co-localisation in human coronary macrophages



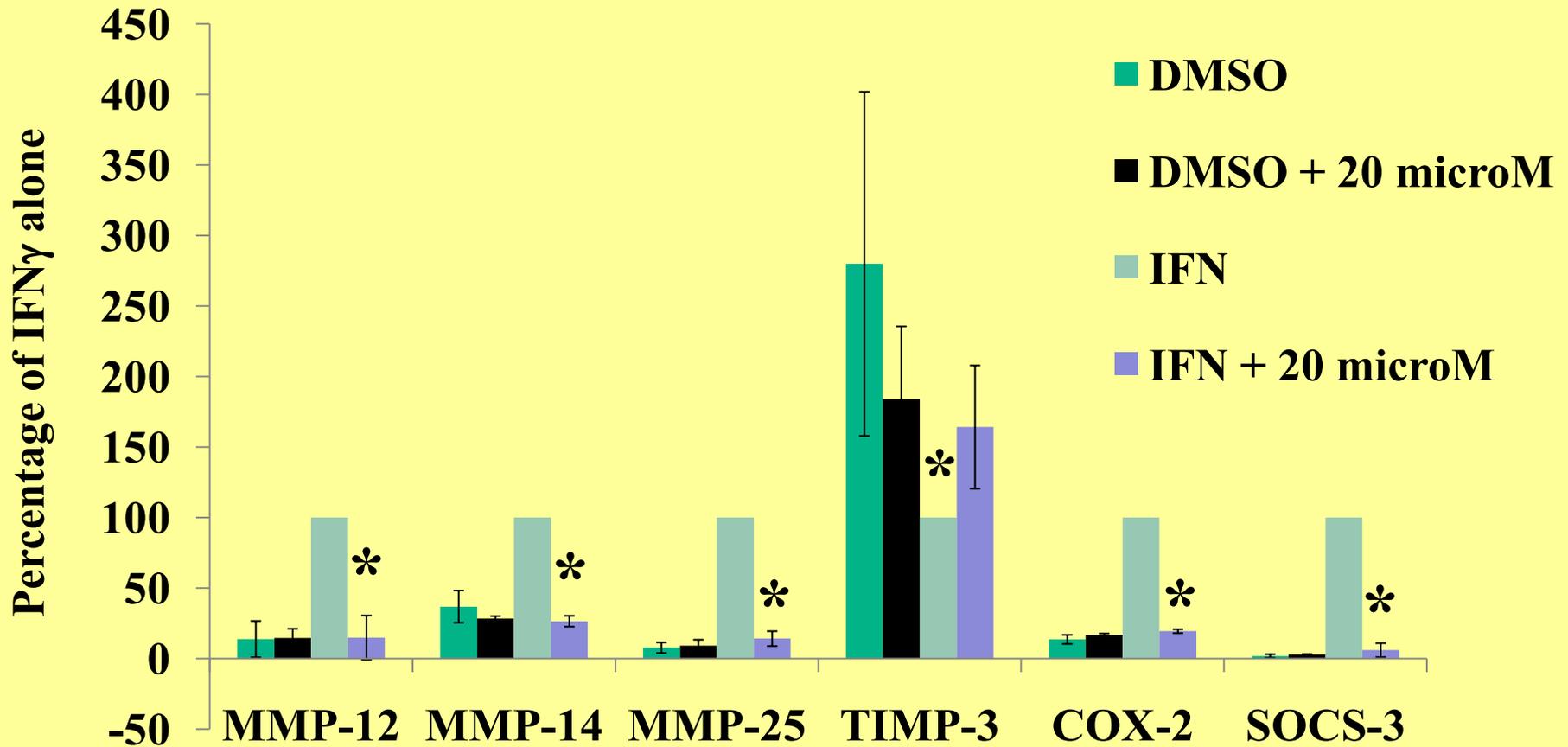
MMP-14 and NF- κ B



Role of TLR2



Effect of JAK-2 inhibitor on IFN γ



Conclusions 1

- M1 genes are regulated through NF κ B in vitro and co-localize in vivo
- A subgroup may co-localize with Stat-1P (to be shown)

Alternatively-activated (M2) phenotypes

Old name	M2a	M2b	M2c	Fibrocyte
Proposed name	MIL4/13	MNHR	MIL10	MTGFβ
Receptor	IL-4RF	GCR, PPAR, Nur77	IL10R	TGFβRI/II
TFs	STAT-6, KLF4	NHR	STAT-3	SMADs2/3 FosB
Markers	CD206, COX-1, maf-1	CD163	CD163	Collagen, fibronectin
Proposed marker	Nuclear STAT6-P			Nuclear SMAD2-P

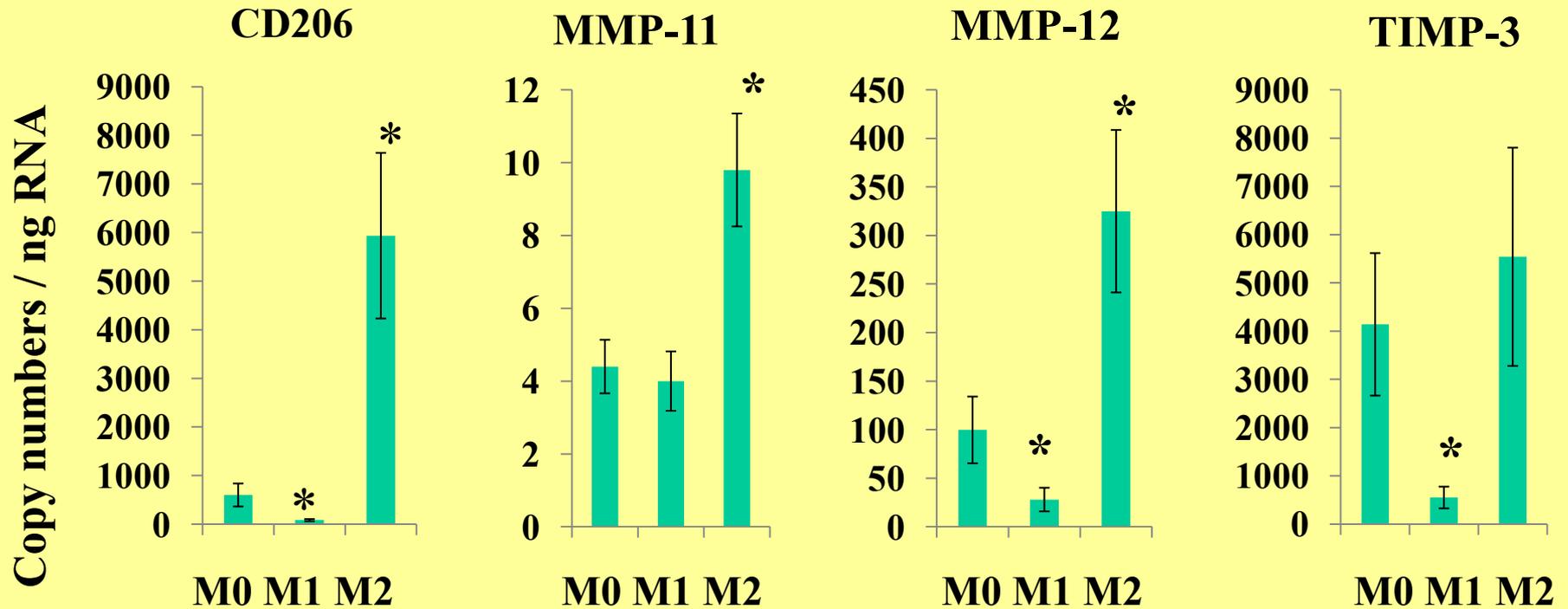
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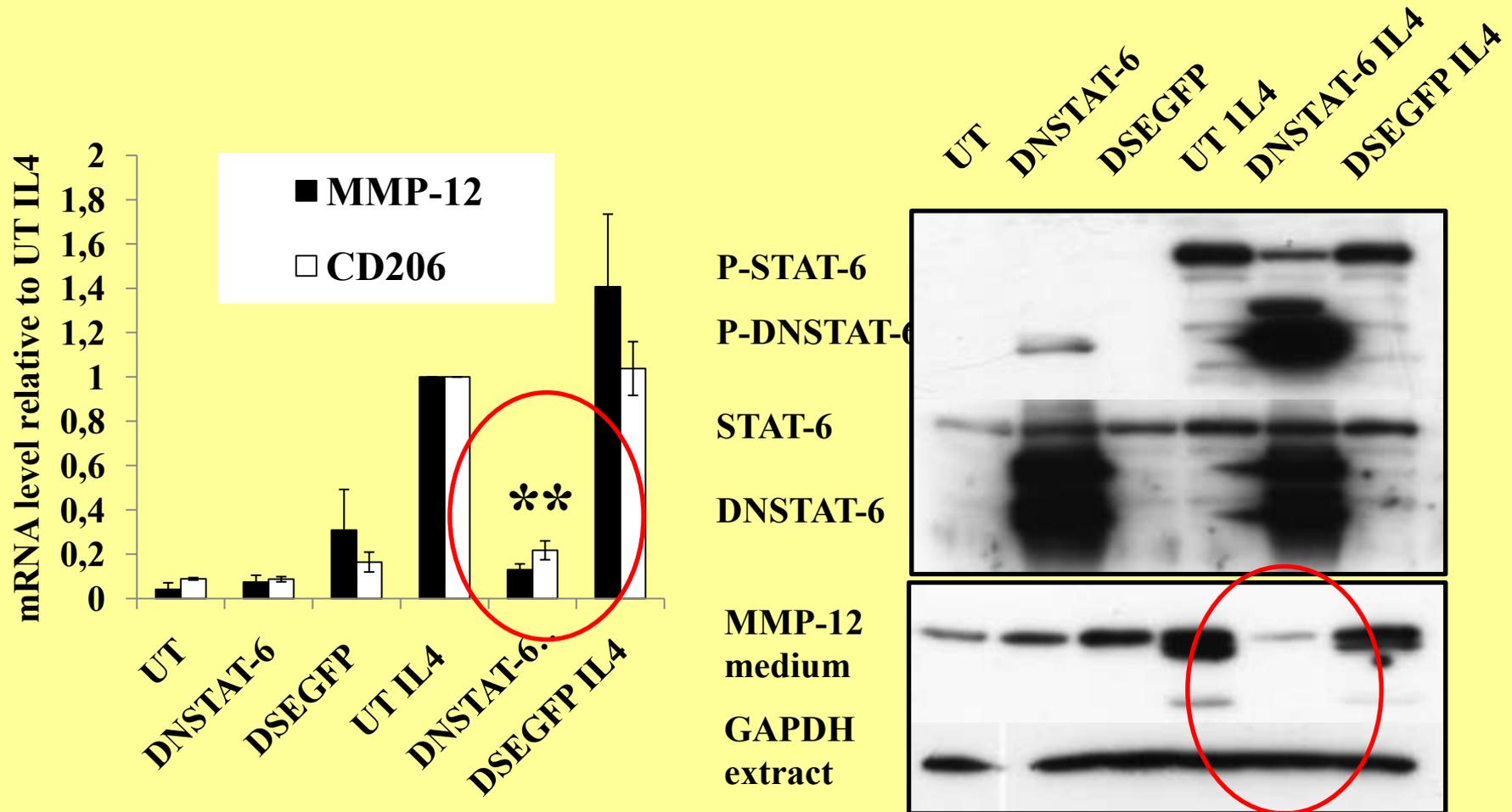
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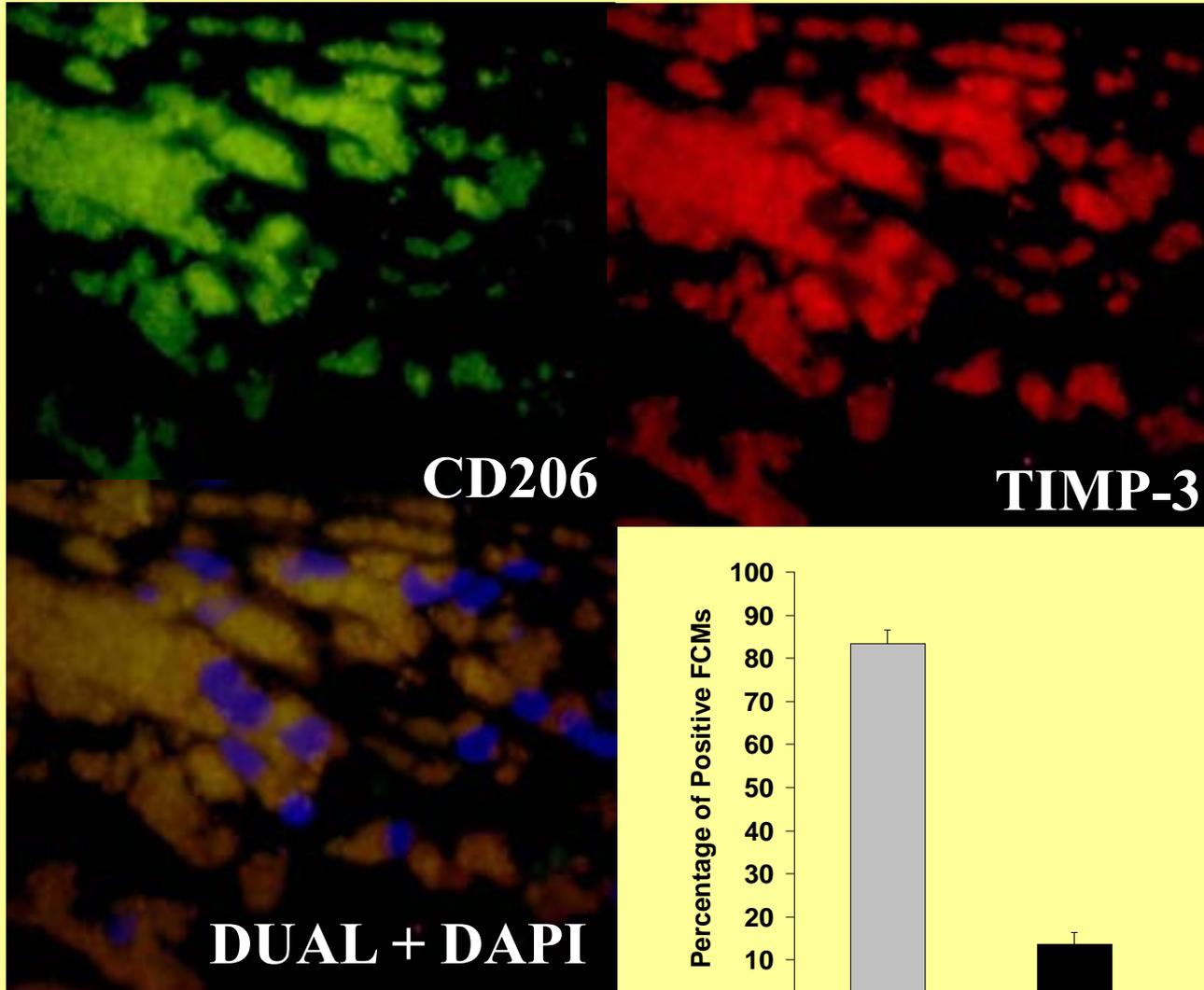
Foam cells stimulated by IL-4 (M2)



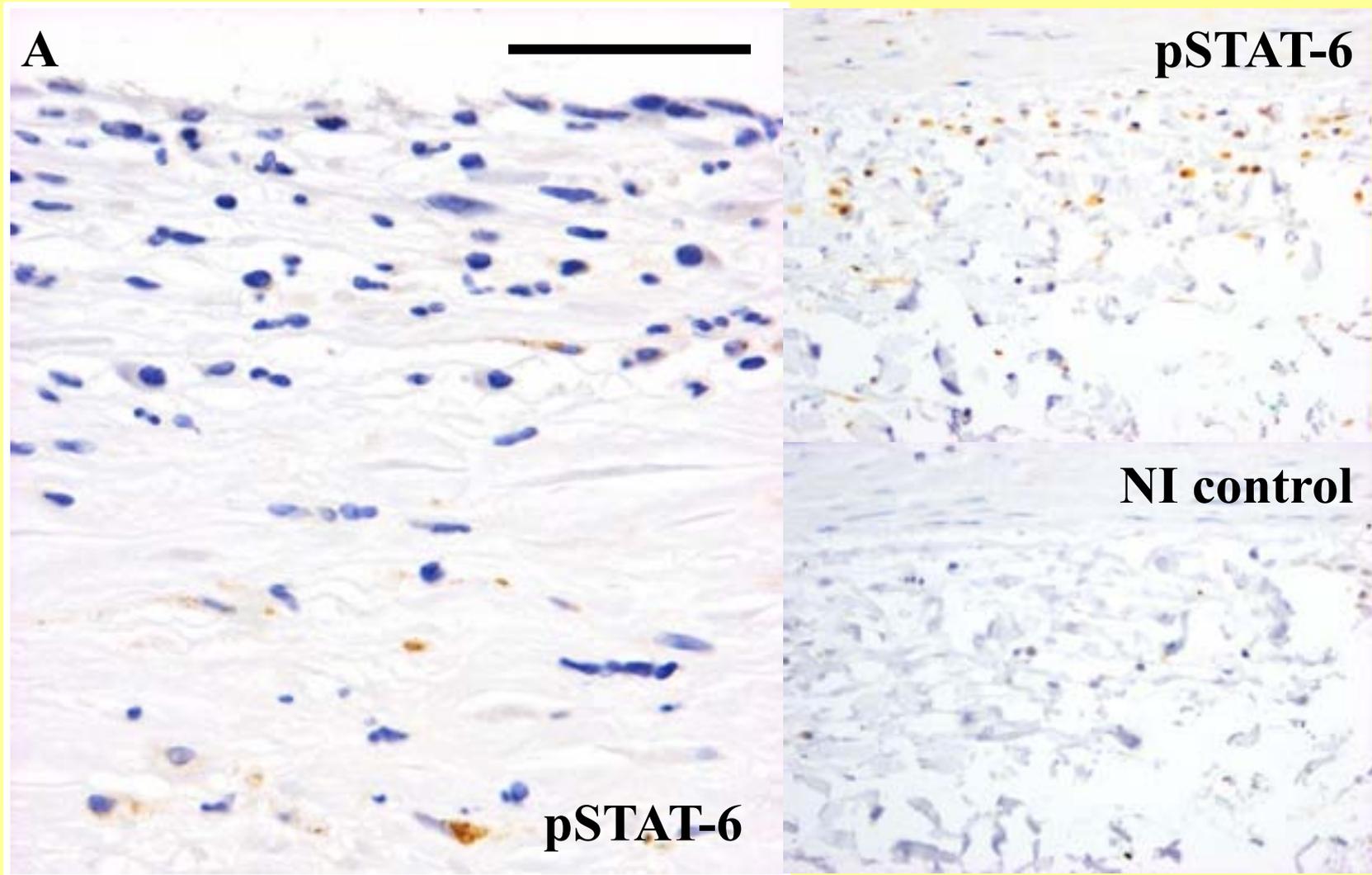
IL-4 up-regulates MMP-12 via STAT6



TIMP-3 and CD206



Is IL4/stat6 up-regulating CD206 in plaques?



Genomic studies on hypoxic macrophages

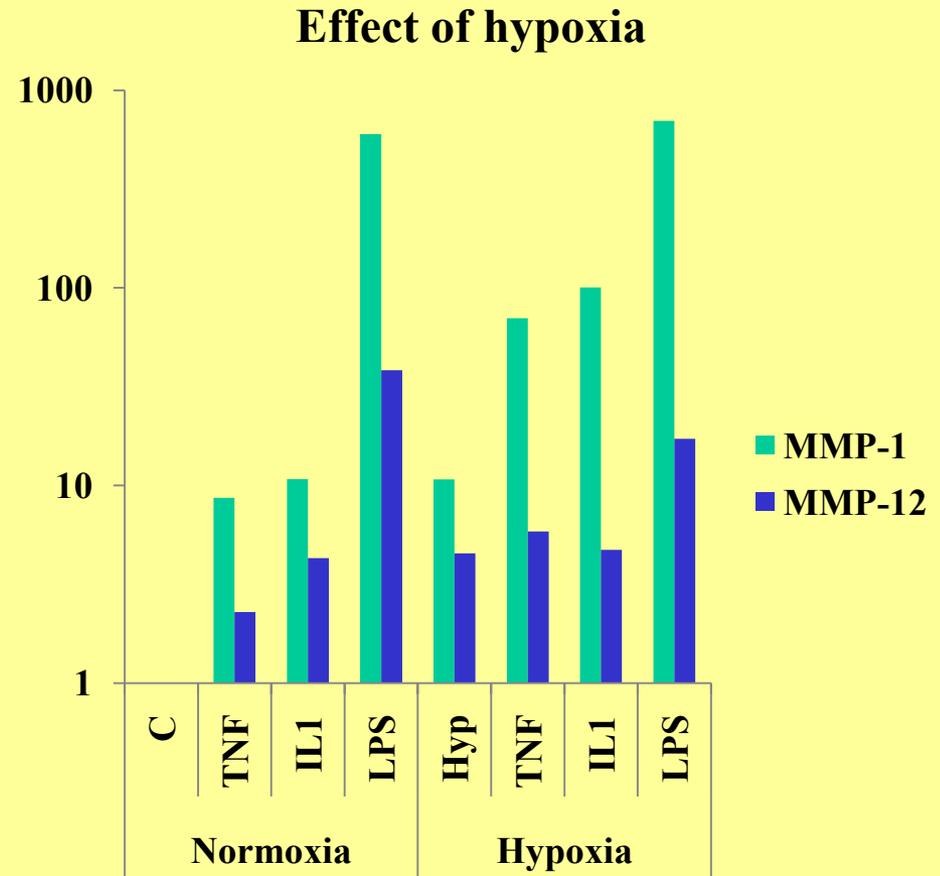
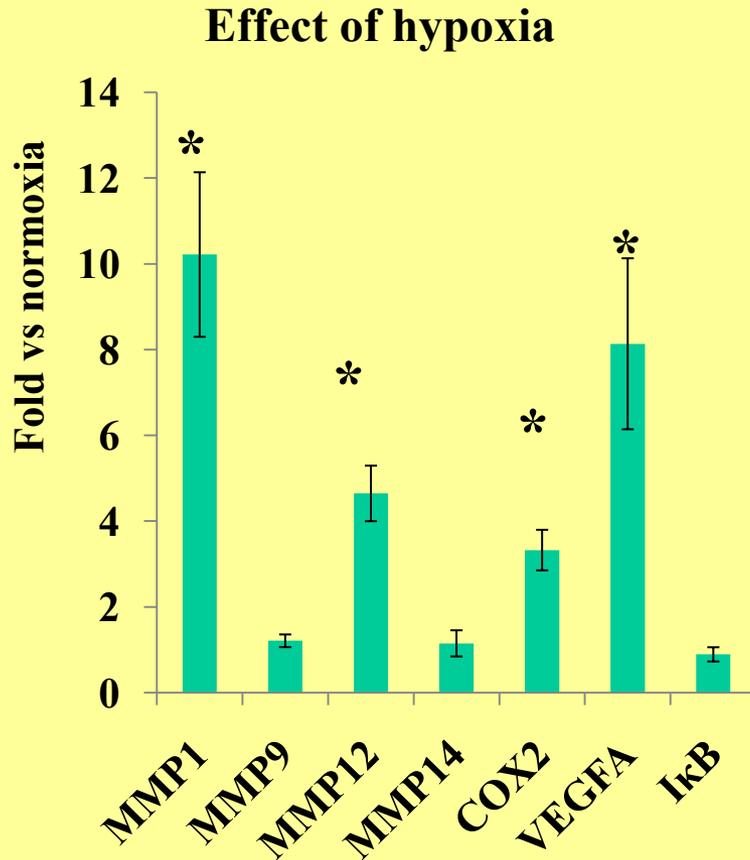
<https://www.nextbio.com/b/study/cor.nb?id=12481#corP=13%2C10%2C99.79840721742082>

Gene	fold change
SERPINB2	699.5
ANKRD1	343.5
IL1A	212.9
IL1B	148.8
LRRC50	88.6
CA12	84.1
CXCL5	69.5
MMP1	67.6
....	
MMP10	35.8
MMP3	11.6
MMP12	3.93

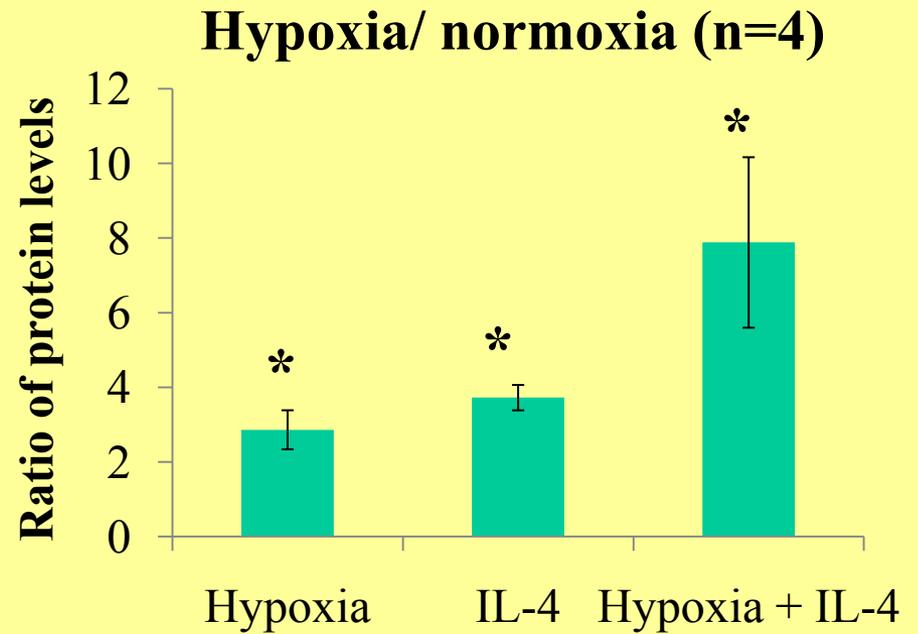
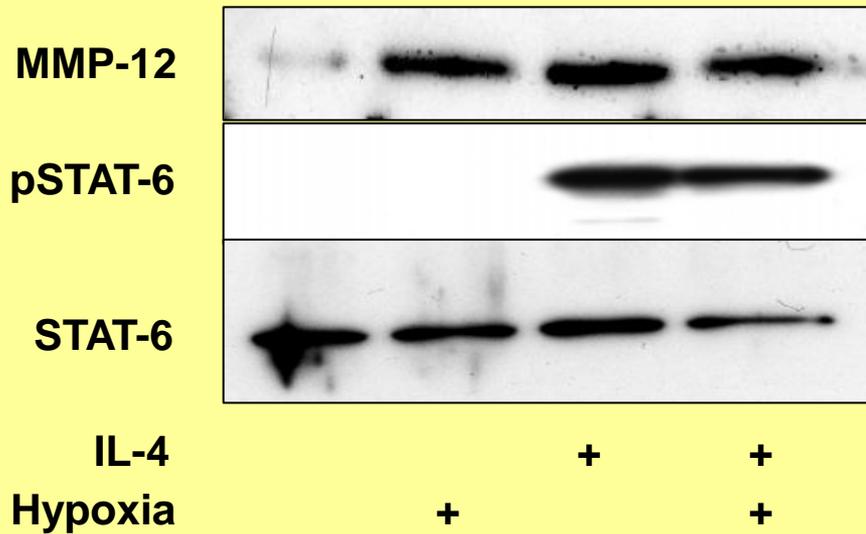
<https://www.nextbio.com/b/study/cor.nb?id=8415>

Gene	fold change
DDIT4	104.1
ADM	52.2
SLC2A3	30.7
TSPAN6	27.7
NR4A3	19.2
MT1X	18.7
NEFL	15.6
GSTA2	15
RGS2	14.1
IL8	13.9
....	
MMP12	3.22

Hypoxia and MMP mRNAs *in vitro*



Hypoxia induced MMP-12 protein independent of Stat-6



Phenotypes related to oxidation

Name	Mox	Mhem	Mhypox
Receptor	ScavengerRs	CD163	
Signal 1	ROS	ROS	HIF1/2 α , ERKs, IKK2, PI3K, wnts
TFs	Nrf2, low NF-κB	Nrf2, ATF-1, low NF-κB	HIF1/2α, NF- κ B, TCF/Lef
Markers	HO-1	HO-1, CD163, Low MHCII	VEGF, Glut-1
Proposed marker	?	?	Nuclear HIF1 α

14

15

16

Conclusions 2

- ‘M2’ genes are regulated through Stat-6 in vitro but do not necessarily co-localize in vivo
- There are a lot more definable phenotypes than M1 and M2
- Follow the pathways to greater wisdom!

Thanks

Buket Reel Graciela Newby



David Huang

